

Comparison of birth weights in patients randomly assigned to fresh or frozen-thawed embryo transfer

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Objective: To estimate birth weight differences between patients randomized to fresh or thawed ET.

Design: Post hoc analysis of results from two similar randomized trials.

Setting: Private fertility center.

Patient(s): One hundred thirty-four first-time IVF patients, ages 18–40 years at oocyte retrieval, who had live birth.

Intervention(s): Patients were randomly assigned to have either fresh blastocyst transfer or all bipronuclear oocytes frozen followed by thaw, extended culture, and blastocyst transfer in a subsequent cycle. Preimplantation genetic screening was not allowed.

Main Outcome Measure(s): Mean birth weight.

Result(s): After allowing for the contributions of multiple significant variables (gestational age at birth, the presence of a vanished twin, number of infants delivered) in multiple linear regression, the adjusted mean birth weight was 166 g (95% confidence interval, 43–290 g) lower after fresh blastocyst transfer when compared with transfer of blastocysts derived from thawed bipronuclear oocytes.

Conclusion(s): Birth weights are lower in cycles with fresh blastocyst transfer after controlled ovarian stimulation than in transfers of frozen-thawed embryos in the absence of ovarian stimulation. This finding confirms similar results reported in many retrospective studies.

Clinical Trial Registration Numbers: NCT00963625 and NCT00963079. (Fertil Steril® 2016;106:317–21. ©2016 by American Society for Reproductive Medicine.)

Key Words: Assisted reproduction, ovarian stimulation, embryo cryopreservation, birth weight

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Controlled ovarian stimulation (COS) with gonadotropins is routinely used to obtain multiple oocytes to increase success rates of IVF as compared with natural ovulation cycles. However, COS and its associated supraphysiologic hormone levels may result in a suboptimal uterine environment for embryo implantation and growth.

COS exposure is associated with altered endometrial development when compared with natural cycles. Developmental differences associated with COS

exposure include histologic advancement, premature down-regulation of the P receptor, an abbreviated luteal phase, glandular-stromal dyssynchrony, genomic dysregulation, altered leukocyte localization and activation, premature nucleolar channel formation, advanced angiogenesis, increased blood vessel density, and reduced endometrial blood flow (1–10). The degree of histologic advancement correlates with IVF outcome and is increased when P levels are prematurely elevated (1, 2, 11–13). One randomized trial

found that embryos transferred in fresh autologous cycles with COS exposure are less likely to implant than their frozen-thawed counterparts transferred in cycles without COS exposure, suggesting a possible effect of reduced endometrial receptivity after ovarian stimulation (14).

It has been repeatedly observed in registry studies and meta-analyses that infants resulting from fresh autologous ET have reduced birth weight, increased risk of low birth weight, and other perinatal risks associated with birth weight when compared with infants resulting from the transfer of frozen-thawed embryos (15–29). For example, the reported birth weight differences, consistently greater with frozen embryos than with fresh, are 167–250 g in Denmark (16–18), 134 g in Finland (19), 133 g in combined

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Nordic registries (20), 156 g in the United States (21), 91–100 g in Japan (22, 23), 145 g in Australia and New Zealand (24), and 80 g among singletons in Latin America (25). Multiple retrospective clinical studies have also found greater birth weight with frozen-thawed ET (FET) than with fresh transfer, with reported birth weight differences ranging from 50 to 218 g (26–29).

One potential cause of birth weight differences is a suboptimal uterine environment after COS exposure (21). Alternatively, cryopreservation might alter birth weight through embryonic effects. However, the contrasting patterns in birth weight effects after autologous fresh, autologous FET, fresh cycles of oocyte donation, and FET cycles using embryos derived from donor oocytes have been used to examine the potential for embryonic or uterine effects of cryopreservation. An American registry study reported no significant difference between the incidence of low birth weight after fresh donor and donor FET cycles, while finding increased incidence of low birth weight after autologous fresh transfer when compared with autologous FET (21). Furthermore, a retrospective cohort study compared births from fresh donor cycles and donor FET cycles in a set of recipients with at least one live birth from each transfer type and found no significant differences in birth weight or perinatal outcomes (30). Each of these findings in oocyte donation cycles contradicts the hypothesis of a significant embryonic effect of cryopreservation on birth weight.

However, retrospective studies, including registry studies, have inherent potential confounding through lack of randomization, and registry studies typically lack detail regarding treatment protocols, such as embryo culture conditions, cryopreservation technique, endometrial preparation, luteal support, and COS protocol, among other potentially relevant parameters. For example, FET cycles have frequently used supernumerary embryos after the morphologically best (primary) embryos were transferred fresh, potentially creating confounding biases in embryo selection and patient selection in retrospective and registry studies. The multiple opportunities for confounding may cloud interpretation of results.

The current study is a post hoc analysis of two concurrent randomized trials, one in normal responders (14) and the other in high responders (31), in which patients were randomly assigned to fresh ET or FET at a single center. The two studies used identical staff, culture conditions, COS protocols and medications, cryopreservation methods, embryo selection and transfer techniques, and luteal support. The inclusion criteria were similar (first-time IVF patients, 18–40 years of age, day 3 FSH <10 IU/L, no preimplantation genetic testing) and differed only in the number of antral follicles so that the normal-responder study (14) specified 8 to 15 antral follicles and the high-responder study (31) specified >15 antral follicles. The purpose of the difference in antral follicle count was so that the normal-responder protocol could specify an ovulatory trigger of hCG alone, while, for safety reasons, the high-responder study specified a trigger of low-dose hCG in combination with GnRH agonist. However, both of these trigger types were allowed in both studies, based on physician discretion. Therefore the two studies form a continuum of

subjects with normal to high ovarian response treated under consistent protocols and laboratory conditions.

Both studies used only primary embryos (no supernumerary embryos) regardless of whether the transfers were fresh or frozen-thawed. The transferred embryos in the FET arms were blastocysts derived from thawed bipronuclear oocytes, so that the morphologically best blastocysts in each study arm were selected for transfer.

A post hoc analysis of these two studies will therefore be useful in examining birth weight differences, if any, without potential confounding from unknown methodological differences or nonrandomized treatment assignment.

MATERIALS AND METHODS

The two randomized trials were performed from 2007 through 2010 and included 259 subjects randomized 1:1 to cohort cryopreservation or else fresh embryo culture at a single infertility center. The two trials were registered on [ClinicalTrials.gov](https://clinicaltrials.gov) with trial numbers NCT00963625 (normal-responder study) and NCT00963079 (high-responder study). Both studies were Institutional Review Board approved and independently monitored. Each study specified an interim stopping point and stopping criteria. The normal-responder study was stopped at that point because the stopping criteria were met (a difference in clinical pregnancy rate with $P < .03$), while the high-responder study was halted at that same point for safety reasons (the routine transfer of two blastocysts in all patients could no longer be supported owing to excessive twins).

Inclusion criteria specified women age 18–40 years seeking their first IVF treatment, with 8–15 antral follicles in the normal-responder study and 16 or more antral follicles in the high-responder study. Cycle day 3 FSH ≥ 10 IU/L was exclusionary, as was any use of preimplantation genetic testing. For the current post hoc study of birth weights, only live births are included in the birth weight analyses.

After informed consent, all subjects underwent conventional COS with gonadotropins, including a combination of urinary FSH (Menopur, Ferring Pharmaceuticals Inc.) and recombinant FSH (Follistim, Schering-Plough Inc.) in all cases. Follicular development was ultrasonically monitored at 2- to 3-day intervals. On or about the sixth day of stimulation, GnRH antagonist (ganirelix acetate, Schering-Plough) was initiated in all patients and sustained until the completion of stimulation.

After at least three follicles reached 18 mm in mean diameter, an ovulatory trigger of either hCG alone or else a reduced dose of hCG in combination with 4 mg GnRH agonist (leuprolide acetate) was administered to promote final oocyte maturation. The high-responder study protocol specified a dual trigger, and the normal-responder study protocol specified a trigger of hCG alone, but variation in the triggers was allowed for safety reasons, and this variation is described in the Results section. For safety reasons, patients who had extreme response to COS (typically more than 30–40 developing follicles) received GnRH agonist trigger alone and were dropped from the studies. Retrieval was scheduled 35–36 hours after trigger. In both studies, randomization was performed by

nonmedical staff drawing identical, unmarked, opaque envelopes immediately after the confirmation that at least one oocyte was collected. All collected oocytes were then inseminated by intracytoplasmic sperm injection.

Subjects randomly assigned to embryo cohort cryopreservation had all of their bipronuclear oocytes frozen by a conventional slow freezing technique described elsewhere in detail (14). In a later menstrual cycle, their entire cohorts of frozen bipronuclear oocytes were thawed at room temperature for 20 minutes in Embryo Thaw Media (Irvine Scientific).

In all cases, fresh or thawed bipronuclear oocytes were cultured to the blastocyst stage in sequential media (Quinn's Advantage Protein Plus Cleavage Media and Quinn's Advantage Protein Plus Blastocyst Media; Sage) before the best two blastocysts (whenever available) were morphologically selected for ultrasound-guided transfer. Patients receiving thawed ET were down-regulated with GnRH agonist (leuprolide acetate), received 10–14 days of oral E₂ (Estrace, 6.0 mg daily) and E₂ patches as needed to achieve an endometrial thickness ≥ 8 mm before P start, and initiated daily P injections (typically 100 mg) on the day before bipronuclear oocyte thaw. Patients randomized to fresh transfer initiated daily P injections 1–2 days after retrieval and E₂ supplements as needed. In all subjects, P and E₂ supplements were continued to sustain serum levels of at least 15 ng/mL and 200 pg/mL, respectively, until a negative pregnancy test, a pregnancy loss, or until rising serum levels demonstrated adequate placental production at approximately 10 weeks' gestation.

Birth information, including dates and weights, was reported by patients or obtained from obstetric records. A vanished twin was identified by the number of ultrasonically observed fetal hearts exceeding the number delivered. Ultrasounds were performed at this center.

Birth weight was the main outcome measure for this post hoc analysis. Wilcoxon's test was used to compare numeric measures, and Fisher's exact test was used to compare nominal outcome measures. All tests were two-tailed, and $P < .05$ was considered statistically significant. Linear regression models (more technically, analysis of covariance, as numeric and dichotomous nominal variables were among the available independent variables) were developed using forward selection ($P < .25$ to enter) followed by backward elimination ($P < .05$ to remain). For regression analysis, the available variables were transfer type (fresh vs. FET), maternal parameters (age at retrieval, weight, height, body mass index, tobacco use, antral follicle count), number transferred, presence of a vanished twin, and number delivered. All analyses were performed with JMP version 7 (SAS Inc.).

RESULTS

The two combined randomized trials included 130 subjects randomized to embryo cohort cryopreservation and 129 randomized to fresh embryo culture. Of these, 99 had FET, and 105 had fresh ET. Live birth was achieved in 74 out of 99 FETs (74.7%) and in 60 of 105 fresh transfers (57.1%), for a total of 134 live births. The live-birth rate per transfer was significantly greater with FET than with fresh transfer ($P = .0119$). The relative risk of failure to

achieve live birth in fresh transfer compared with FET was 1.69 (95% confidence interval [CI], 1.13–2.54). The live-birth rates per randomized subject were 56.9% in subjects randomized to cohort cryopreservation and 45.5% in those randomized to fresh embryo culture ($P = .1065$), and the relative risk of failure to achieve live birth among patients randomized to fresh culture compared with those randomized to cohort cryopreservation was 1.24 (95% CI, 0.96–1.60).

Table 1 describes and compares cycles resulting in live birth with fresh ETs and FETs. No significant differences were observed in maternal age at retrieval, maternal weight, maternal height, maternal body mass index, tobacco use, antral follicle count, duration of stimulation, total FSH dose, serum E₂ level on the day of trigger, trigger medication, number of collected oocytes, number of mature oocytes, frequency of vanished twins, number of delivered infants, or the number of singletons, twins, or triplets. There were no

TABLE 1

Univariate comparisons of patient parameters, cycle characteristics, and outcomes.

Variable	Fresh transfer	FET	P value
Births, n	60	74	
Maternal age, y	31.3 $\pm$ 3.6	31.3 $\pm$ 4.0	.8241
Maternal weight, kg	69.5 $\pm$ 16.3	70.3 $\pm$ 17.6	.8086
Maternal height, cm	165 $\pm$ 8	164 $\pm$ 7	.1810
Maternal BMI, kg/m ²	25.3 $\pm$ 5.3	26.1 $\pm$ 5.9	.5647
Maternal tobacco user	12/60	18/74	.6775
Antral follicles, n	18.8 $\pm$ 9.3	18.3 $\pm$ 7.3	.8986
Total FSH dose, IU	2,528 $\pm$ 745	2,649 $\pm$ 828	.3252
Stimulation period, d	10.2 $\pm$ 1.3	10.3 $\pm$ 1.2	.6207
E ₂ , day of trigger, pg/mL	4,081 $\pm$ 1,627	4,000 $\pm$ 2,277	.3361
Type of trigger, n			
hCG only	16/60	29/74	.1442
Dual trigger	44/60	45/74	.1442
Oocytes collected, n	17.9 $\pm$ 8.1	17.7 $\pm$ 8.3	.7555
Mature oocytes collected, n	14.0 $\pm$ 6.0	13.9 $\pm$ 6.6	.7132
Blastocysts transferred, n	1.92 $\pm$ 0.28	1.88 $\pm$ 0.33	.4729
Vanished twin, n	7/60	10/74	.7996
Delivered, n	1.42 $\pm$ 0.53	1.46 $\pm$ 0.53	.6123
Mean birth weight, kg	2.739 $\pm$ 0.655	2.873 $\pm$ 0.775	.2632
Gestational age at birth, d	258.5 $\pm$ 20.3	258.0 $\pm$ 22.6	.9750
Singleton births	36	41	
Mean birth weight, kg	3.076 $\pm$ 0.511	3.242 $\pm$ 0.701	.1572
Gestational age at birth, d	268.3 $\pm$ 12.7	268.3 $\pm$ 13.4	.9593
Twin births	23	32	
Mean birth weight, kg	2.246 $\pm$ 0.516	2.422 $\pm$ 0.611	.1143
Gestational age at birth, d	244.0 $\pm$ 21.3	245.5 $\pm$ 25.6	.3059
Triplet births, n	1	1	
Mean birth weight, kg	1.928	2.165	NA
Gestational age at birth, d	242.6	234.5	NA

Shapiro. Birth weight comparison. *Fertil Steril* 2016.

significant differences in mean birth weights or gestational age at birth between fresh transfer and FET, nor when broken out by the number delivered (Table 1).

Because of the potential for multiple confounding variables affecting birth weight, most notably gestational age at birth, multiple linear regression was used to investigate in greater detail whether any of the available variables might explain the variation in mean birth weight.

After forward stepwise selection and backward elimination, multiple linear regression identified the following significant predictors of mean birth weight: gestational age at birth ($P < .0001$), number of delivered infants ($P = .0002$), vanished twin ($P = .0060$), and FET ($P = .0094$). Each additional day of gestational age was estimated to increase mean birth weight by 24.8 g (95% CI, 21.3–28.3 g), each increment in the number delivered was estimated to reduce birth weight by 29.1 g (95% CI, 14.2–44.0 g), a vanished twin was estimated to reduce birth weight by 282 g (95% CI, 84–480 g), and FET increased birth weight by 166 g (95% CI, 43–290 g) when compared with fresh transfer.

DISCUSSION

Birth weight differences were not significant in the univariate analyses in Table 1. While mean birth weights were consistently 166–237 g greater in FET than in fresh transfer across each number delivered, none of these comparisons were statistically significant. A multivariate procedure, multiple linear regression, was employed to discern the contributions of each significant variable.

This is the first report comparing birth weights in patients randomly assigned to fresh or FET. Many prior studies have investigated the potential effects of COS and cryopreservation on birth weight. These have all been retrospective studies, either of national registries or of results at individual centers. Studies of national registries have very large sample sizes but lack detail regarding treatment assignment and medical methodology, and retrospective studies generally confer substantial opportunities for confounding effects through lack of randomization. For example, a common weakness in retrospective studies comparing fresh transfer and FET is the potential that the former includes only primary embryos, while the latter is mainly supernumerary embryos after the morphologically best embryos were transferred fresh.

The current finding suggests there were no substantial confounding uncontrolled variables causing the birth weight differences in the prior studies. The current study, although small in sample size, has the advantages of known and detailed methodology and randomized treatment assignment. For example, all patients in both groups had the same embryo culture conditions and had only their primary (morphologically best) embryos selected for transfer, and none had genetic screening of embryos.

The results were consistent with the conclusions of many prior retrospective studies (16–29), suggesting those prior findings were not due to confounded uncontrolled variables. Specifically, the current study found 166 g (95% CI) greater birth weight after FET when compared with fresh

transfer, a value very near the center of the range of 80–250 g difference reported in 10 registry studies (16–25) and very close to the 156 g reported value from analysis of a U.S. registry (21).

The two randomized trials combined here had very similar inclusion criteria and methodology. The only methodological difference in the protocols was that the high-responder study specified a dual trigger as the primary therapy to reduce ovarian hyperstimulation syndrome risk, while the normal-responder study specified hCG alone. However, both trigger types were allowed in both studies per physician judgment, and the normal-responder study also included 20 births that followed dual triggers after a greater than expected ovarian response, while the high-responder study included one live birth that followed an hCG trigger in a patient with lower ovarian response than expected. Therefore, there was no clear distinction in methodology between the two trials, and their combined data set is actually a continuum of subjects with at least eight antral follicles. The proportion of use of each trigger did not differ significantly between the fresh and FET groups, and the randomization always occurred immediately after oocyte retrieval so that the physician's trigger choice could not be influenced by treatment assignment.

The current study cannot resolve whether the observed birth weight difference resulted from an embryonic effect of cryopreservation or an endometrial effect of COS. However, comparisons of birth weights in cycles using embryos derived from donor oocytes have reported no difference in birth weight or in the incidence of low birth weight between fresh transfers and FET, contradicting the hypothesis of an embryonic effect of cryopreservation on birth weight in the absence of uterine COS exposure (21, 30).

The cryopreservation protocol used here was conventional slow freezing of bipronuclear oocytes, followed by thaw, extended culture, and blastocyst transfer. This method is associated with reduced blastocyst yield when compared with extended culture from fresh oocytes (32) and has been largely replaced by blastocyst vitrification (33). However, the resulting blastocysts from this technique have implantation potential comparable to that of vitrified-thawed blastocysts, and the artificial endometrial preparation is similar in both techniques (33). Should there prove to be any embryonic effects of cryopreservation on birth weight, then the results here might be method specific and not specifically relevant to blastocyst vitrification. However, as discussed above, current evidence suggests the frequently observed birth weight differences between fresh and thawed ETs result from uterine effects, and therefore the results presented here and in the cited registry studies should be relevant to other cryopreservation protocols.

The live-birth rate per transfer was significantly greater with FET than with fresh transfer, suggesting inferior endometrial receptivity in fresh transfer after COS exposure as concluded before (14). The causal mechanisms behind the altered endometrial receptivity may be related to those affecting birth weight. However, the live-birth rate per randomized subject did not differ significantly. Therefore more evidence is needed to determine whether cohort

cryopreservation improves the chance of live birth. However, the birth weight differences reported here and in the many cited registry studies remain a significant factor to consider.

REFERENCES

- Nikas G, Develioglu OH, Toner JP, Jones HW Jr. Endometrial pinopodes indicate a shift in the window of receptivity in IVF cycles. *Hum Reprod* 1999;14:787–92.
- Kolibianakis E, Bourgain C, Albano C, Osmanagaoglu K, Smits J, Van Steirteghem A, et al. Effect of ovarian stimulation with recombinant follicle-stimulating hormone, gonadotropin releasing hormone antagonists, and human chorionic gonadotropin on endometrial maturation on the day of oocyte pick-up. *Fertil Steril* 2002;78:1025–9.
- Develioglu OH, Hsiu JG, Nikas G, Toner JP, Oehninger S, Jones HW Jr. Endometrial estrogen and progesterone receptor and pinopode expression in stimulated cycles of oocyte donors. *Fertil Steril* 1999;71:1040–7.
- Mirkin S, Nikas G, Hsiu JG, Diaz J, Oehninger S. Gene expression profiles and structural/functional features of the peri-implantation endometrium in natural and gonadotropin-stimulated cycles. *J Clin Endocrinol Metab* 2004;89:5742–52.
- Horcajadas JA, Diaz-Gimeno P, Pellicer A, Simon C. Uterine receptivity and the ramifications of ovarian stimulation on endometrial function. *Semin Reprod Med* 2007;25:454–60.
- Haouzi D, Assou S, Mahmoud K, Tondeur S, Reme T, Hedon B, et al. Gene expression profile of human endometrial receptivity: comparison between natural and stimulated cycles for the same patients. *Hum Reprod* 2009;24:1436–45.
- Evans J, Hannan NJ, Hincks C, Rombauts LJ, Salamonsen LA. Defective soil for a fertile seed? Altered endometrial development is detrimental to pregnancy success. *PLoS One* 2012;7:e53098.
- Zapantis G, Szymiga MJ, Rybak EA, Meier UT. Premature formation of nucleolar channel systems indicates advanced endometrial maturation following controlled ovarian hyperstimulation. *Hum Reprod* 2013;28:3292–300.
- Lee YL, Liu Y, Ng PY, Lee KF, Au CL, Ng EH, et al. Aberrant expression of angiopoietins-1 and -2 and vascular endothelial growth factor-A in peri-implantation endometrium after gonadotrophin stimulation. *Hum Reprod* 2008;23:894–903.
- Ng EH, Chan CC, Tang OS, Yeung WS, Ho PC. Comparison of endometrial and subendometrial blood flow measured by three-dimensional power Doppler ultrasound between stimulated and natural cycles in the same patients. *Hum Reprod* 2004;19:2385–90.
- Venetis CA, Kolibianakis EM, Bosdou JK, Tarlatzis BC. Progesterone elevation and probability of pregnancy after IVF: a systematic review and meta-analysis of over 60,000 cycles. *Hum Reprod Update* 2013;19:433–57.
- Venetis CA, Kolibianakis EM, Bosdou JK, Lainas GT, Sfountouris IA, Tarlatzis BC, et al. Estimating the net effect of progesterone elevation on the day of hCG on live birth rates after IVF: a cohort analysis of 3296 IVF cycles. *Hum Reprod* 2015;30:684–91.
- Chetkowski RJ, Kiltz RJ, Salyer WR. In premature luteinization, progesterone induces secretory transformation of the endometrium without impairment of embryo viability. *Fertil Steril* 1997;68:292–7.
- Shapiro BS, Daneshmand ST, Garner FC, Aguirre M, Hudson C, Thomas S. Evidence of impaired endometrial receptivity after ovarian stimulation for in vitro fertilization: a prospective randomized trial comparing fresh and frozen-thawed embryo transfer in normal responders. *Fertil Steril* 2011;96:344–8.
- Maheshwari A, Pandey S, Shetty A, Hamilton M, Bhattacharya S. Obstetric and perinatal outcomes in singleton pregnancies resulting from the transfer of frozen thawed versus fresh embryos generated through in vitro fertilization treatment: a systematic review and meta-analysis. *Fertil Steril* 2012;98:368–77.
- Pinborg A, Henningsen AA, Loft A, Malchau SS, Forman J, Andersen AN. Large baby syndrome in singletons born after frozen embryo transfer (FET): is it due to maternal factors or the cryotechnique? *Hum Reprod* 2014;29:618–27.
- Pinborg A, Loft A, Aaris Henningsen AK, Rasmussen S, Andersen AN. Infant outcome of 957 singletons born after frozen embryo replacement: the Danish National Cohort Study 1995–2006. *Fertil Steril* 2010;94:132–7.
- Henningsen AK, Pinborg A, Lidegaard O, Vestergaard C, Forman JL, Andersen AN. Perinatal outcome of singleton siblings born after assisted reproductive technology and spontaneous conception: Danish national sibling-cohort study. *Fertil Steril* 2011;95:959–63.
- Pelkonen S, Koivunen R, Gissler M, Nuojua-Huttunen S, Suikkari AM, Hyden-Granskog C, et al. Perinatal outcome of children born after frozen and fresh embryo transfer: the Finnish cohort study 1995–2006. *Hum Reprod* 2010;25:914–23.
- Wennerholm UB, Henningsen AK, Romundstad LB, Bergh C, Pinborg A, Skjaerven R, et al. Perinatal outcomes of children born after frozen-thawed embryo transfer: a Nordic cohort study from the CoNARTaS group. *Hum Reprod* 2013;28:2545–53.
- Kalra SK, Ratcliffe SJ, Coutifaris C, Molinaro T, Barnhart KT. Ovarian stimulation and low birth weight in newborns conceived through in vitro fertilization. *Obstet Gynecol* 2011;118:863–71.
- Ishihara O, Araki R, Kuwahara A, Itakura A, Saito H, Adamson GD. Impact of frozen-thawed single-blastocyst transfer on maternal and neonatal outcome: an analysis of 277,042 single-embryo transfer cycles from 2008 to 2010 in Japan. *Fertil Steril* 2014;101:128–33.
- Nakashima A, Araki R, Tani H, Ishihara O, Kuwahara A, Irahara M, et al. Implications of assisted reproductive technologies on term singleton birth weight: an analysis of 25,777 children in the national assisted reproduction registry of Japan. *Fertil Steril* 2013;99:450–5.
- Li Z, Wang YA, Ledger W, Edgar DH, Sullivan EA. Clinical outcomes following cryopreservation of blastocysts by vitrification or slow freezing: a population-based cohort study. *Hum Reprod* 2014;29:2794–801.
- Schwarze JE, Crosby JA, Zegers-Hochschild F. Effect of embryo freezing on perinatal outcome after assisted reproduction techniques: lessons from the Latin American Registry of Assisted Reproduction. *Reprod Biomed Online* 2015;31:39–43.
- Shih W, Rushford DD, Bourne H, Garrett C, McBain JC, Healy DL, et al. Factors affecting low birthweight after assisted reproduction technology: difference between transfer of fresh and cryopreserved embryos suggests an adverse effect of oocyte collection. *Hum Reprod* 2008;23:1644–53.
- Vergouw CG, Kostelijk EH, Doejaaren E, Hompes PG, Lambalk CB, Schats R. The influence of the type of embryo culture medium on neonatal birthweight after single embryo transfer in IVF. *Hum Reprod* 2012;27:2619–26.
- Hu XL, Feng C, Lin XH, Zhong ZX, Zhu YM, Lv PP, et al. High maternal serum estradiol environment in the first trimester is associated with the increased risk of small-for-gestational-age birth. *J Clin Endocrinol Metab* 2014;99:2217–24.
- Korosec S, Ban Frangez H, Verdenik I, Kladnik U, Kotar V, Virant-Klun I, et al. Singleton pregnancy outcomes after in vitro fertilization with fresh or frozen-thawed embryo transfer and incidence of placenta praevia. *Biomed Res Int* 2014;2014:431797.
- Galliano D, Garrido N, Serra-Serra V, Pellicer A. Difference in birth weight of consecutive sibling singletons is not found in oocyte donation when comparing fresh versus frozen embryo replacements. *Fertil Steril* 2015;104:1411–8.
- Shapiro BS, Daneshmand ST, Garner FC, Aguirre M, Hudson C, Thomas S. Evidence of impaired endometrial receptivity after ovarian stimulation for in vitro fertilization: a prospective randomized trial comparing fresh and frozen-thawed embryo transfers in high responders. *Fertil Steril* 2011;96:516–8.
- Shapiro BS, Daneshmand ST, Garner FC, Aguirre M, Hudson C, Thomas S. Embryo cryopreservation rescues cycles with premature luteinization. *Fertil Steril* 2010;93:636–41.
- Shapiro BS, Daneshmand ST, Garner FC, Aguirre M, Hudson C. Freeze-all at the blastocyst or bipronuclear stage: a randomized clinical trial. *Fertil Steril* 2015;104:1138–44.